## **Natural Product Synthesis**

DOI: 10.1002/ange.200702707

## Enantioselective Synthesis of the Complex Rocaglate (-)-Silvestrol\*\*

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Extracts from dried roots and stems of several species of the plant genus *Aglaia* are the source of the rocaglamides, a unique group of natural products featuring a cyclopenta[b]te-trahydrobenzofuran skeleton.<sup>[1]</sup> The complex rocaglate silvestrol (1) and its epimer 2 (Figure 1) were recently isolated

Figure 1. (-)-Silvestrol and related derivatives.

from the plant Aglaia foveolata by Kinghorn and coworkers. [2,3] The structural assignment of 1 was based on NMR spectroscopy and X-ray diffraction studies of the bis-p-bromobenzoate derivative 3. Compound 1 was shown to exhibit very potent cytotoxic activity (e.g.  $ED_{50} = 1.2 \text{ nm}$  against human lung cancer cells) which is comparable to the activity of the anticancer agent Paclitaxel (Taxol). Mechanism-of-action studies indicate that cytotoxicity induced by silvestrol in human prostate cancer (LNCaP) cells is associated with a block in the cell cycle at the G2/M checkpoint in a manner that is independent of p53.[3] In contrast to other rocaglate derivatives, silvestrol possesses a unique dioxanyloxy group at C6 of the cyclopenta[b]benzofuran core. Although the rocaglamides [1c,d] and the corresponding dioxanyloxy side chain<sup>[1e,f]</sup> of silvestrol have been the subject of synthetic studies, an enantioselective synthesis of silvestrol has not been reported to date. We have chosen the promising anticancer lead compound silvestrol as a synthetic target to further develop applications of our enantioselective photogeneration/cycloaddition of oxido-

pyryliums derived from 3-hydroxyflavones<sup>[4]</sup> and to develop chemistry pertaining to the synthesis and attachment of the structurally unique dioxanyloxy segment.

Our retrosynthetic analysis of silvestrol (1) is illustrated in Figure 2. Compound 1 may be derived from C-O bond formation between 1,4-dioxan-2-ol 4 and hydroxyphenyl rocaglate derivative 5. Compound 5 may be prepared using photocycloaddition methodology developed in our laboratory to access the methyl

**Figure 2.** Retrosynthetic analysis of (-)-silvestrol (1).

rocaglate core by employing protected 3-hydroxyflavone **6** as starting material.<sup>[4]</sup> The unusual dioxanyloxy fragment **4** may be obtained via 1,4-dioxan-2-one precursor **7**, which may be derived from 1,2-dibenzylthreitol derivative **8**. Intermediate **8** is readily obtained from the commercially available reagent D-dimethyl tartrate.<sup>[5]</sup>

The synthesis of the protected 3-hydroxyflavone 6 began with Friedel–Crafts acylation of phloroglucinol (9) using benzyloxyacetyl chloride in the presence of AlCl<sub>3</sub> following a modified procedure (Scheme 1).<sup>[6]</sup> After MOM protection and methylation, the derived aryl ketone 10 was subjected to selective MOM deprotection in the presence of a catalytic amount of iodine in methanol,<sup>[7]</sup> followed by acylation with 4-methoxybenzoyl chloride, to afford phenyl ester 11. Baker–Venkataraman rearrangement of 11 under basic conditions (LiHMDS, THF) yielded diketone 12.<sup>[8]</sup> Treatment of 12 with sodium acetate in acetic acid effected smooth cyclization/dehydration<sup>[9]</sup> with concomitant removal of the MOM ether. Reintroduction of the MOM protecting group and subsequent hydrogenolysis of the benzyl group provided the requisite 3-hydroxyflavone 6.

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[\*\*] Financial support from the National Institutes of Health (J.A.P.,Jr., GM-073855) and from the NCI Canada (J.P., 017099), Bristol-Myers Squibb, and Merck Research Laboratories is gratefully acknowledged. We thank Dr. R. Murray Tait for providing natural silvestrol and Dr. Siva Dandapani, Dr. Jean-Charles Marié, Dr. Aaron Beeler, and Mr. Suwei Dong (Boston University) for helpful discussions.
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Scheme 1. a) Benzyloxyacetyl chloride, AlCl<sub>3</sub>, CH<sub>2</sub>Cl<sub>2</sub>/Et<sub>2</sub>O (1:1), 50 °C, 24 h, 58%; b) MOMCl,  $K_2CO_3$ , acetone, RT, 3 h, 60%; c)  $Me_2SO_4$ ,  $K_2CO_3$ , acetone, 60°C, 6 h, 93%; d)  $I_2$ , MeOH, RT, 4 h, 94%; e) 4-methoxybenzoyl chloride,  $Et_3N$ , DMAP,  $CH_2Cl_2$ , RT, 4 h, 85%; f) LiHMDS (3 equiv), THF, -20°C, 1 h, 88%; g) AcOH, AcONa (2.5 equiv), 100°C, 3 h; h) MOMCl, acetone,  $K_2CO_3$ , RT, 6 h, 70% over two steps; i)  $H_2$ ,  $Pd(OH)_2$ , EtOH/THF (1:1), RT, 45 min, 92%. Bn = benzyl, MOM = methoxymethyl, DMAP = 4-dimethylaminopyridine, HMDS = hexamethyldisilazide.

With compound 6 in hand, we proceeded to evaluate the asymmetric synthesis of methyl rocaglate fragment 5 by employing enantioselective [3+2] photocycloaddition mediated by functionalized TADDOL derivatives (TADDOL =  $\alpha, \alpha, \alpha', \alpha'$ -tetraaryl-2,2-dimethyl-1,3-dioxolan-4,5-dimethanol).[4b] During our previous investigations, we found that both the nature of the aryl substituent and ketal side chain of the TADDOL framework, as well as low-temperature reaction conditions, were crucial factors for high enantioselectivity. Photocycloaddition ( $h\nu > 350 \text{ nm}$ ) of 3-hydroxyflavone 6 and methyl cinnamate (13) in the presence of chiral additive 14, bearing a 1-pyrenyl substituent and cyclooctyl ketal, at -70°C using PhCH<sub>3</sub>/CH<sub>2</sub>Cl<sub>2</sub> (2:1) as solvent led to the formation of cycloadduct 15 as well as its ketol-shift isomer<sup>[4a]</sup> after

purification on SiO<sub>2</sub> (Scheme 2). After an α-ketol rearrangement/hydroxy-directed reduction sequence, [4a] endo-rocaglate derivative 16 was isolated in 57% yield and 71% ee along with the corresponding exo stereoisomer 17. Compound 16 was then subjected to MOM deprotection using TMSBr in CH<sub>2</sub>Cl<sub>2</sub> to afford hydroxyphenyl rocaglate derivative 5. Fortunately, we were able to increase the enantiomeric excess of 5 through recrystallization to afford centrosymmetric racemate crystals<sup>[4b]</sup> and 5 with 87 % ee (75 % recovery) in the mother liquor.

The synthesis of the dioxanyloxy fragment 4 was initiated with (2S,3S)-1,2-di-O-benzylidenethreitol (8), which was readily obtained in four steps from commercially available D-dimethyl tartrate.<sup>[5]</sup> Our first strategy to obtain 1,4-

dioxan-2-one 7 was based on a threestep synthesis starting from 8 involving protection of the primary alcohol, followed by esterification with dimethoxyacetic acid, and final deprotection of the primary alcohol and attempted transacetalization of 18 under acid conditions (e.g. p-TsOH, CSA, K-10 clay) (Scheme 3). However, using this pathway we could not isolate the desired dioxane derivative 7. The major product isolated was characterized as the primary ester derivative obtained from acyl transfer during the deprotection step. A similar approach led us to investigate a one-pot process for functionalization of the 1,2-diol 8 through regioselective alkylation followed by lactonization. We thus envisioned the use of a tin acetal as a reactive intermediate for

**Scheme 2.** a)  $h\nu > 350$  nm,  $CH_2Cl_2/toluene$ , -70 °C, 10 h, 66%; b) MeONa (2.5 equiv), MeOH, 60°C, 30 min, 89%; c) Me<sub>4</sub>NBH(OAc)<sub>3</sub>, MeCN, AcOH, 57% (16), 13% (17); d) TMSBr, CH<sub>2</sub>Cl<sub>2</sub>, 3 h, -78 °C, 84% yield, 87% ee after recrystallization. TMS = trimethylsilyl.

regioselective alkylation. Formation of the O-stannylene acetal<sup>[10]</sup> derived from 8, followed by addition of freshly prepared methyl 2-bromo-2-methoxy acetate, [11] afforded 7 (45%) and its stereoisomer 19 (33%). The overall process 8→7 represents a tandem alkylation/lactonization sequence.<sup>[12]</sup> Attempted epimerization of dioxanyl derivative 19 was unsuccessful and led mostly to decomposition. Finally, DIBAL reduction of 7 produced 1,4-dioxan-2-ol 4 as a mixture of diastereoisomers.[13]

With the two fragments 4 and 5 in hand, we evaluated a series of conditions for their coupling (Scheme 4).<sup>[14]</sup> Utilization of glucosidation methods using fluoride or trichloroacetamidate reagents derived from 4 led to unsatisfactory yields of coupling products and significant decomposition. After

Scheme 3. a) TBSCl, imidazole, Et<sub>3</sub>N, DMF, RT, 6 h; b) dimethoxyacetic acid, DCC, DMAP, THF, RT, 82% over two steps; c) nBu<sub>2</sub>SnO, benzene, reflux 9 h, CH<sub>3</sub>OCHBrCO<sub>2</sub>Me, TBAI, benzene, 70°C, 2 h 45% (7), 33% (18); d) DIBAL-H, toluene, -78°C, 1 h, 83 %. TBS = tert-butyldimethylsilyl, DCC = dicyclohexyl carbodiimide, TBAI = tetra-n-butylammonium iodide, DIBAL-H = diisobutylaluminum hy-

Scheme 4. a) DIAD, F-PPh3, toluene, 4-Å MS, RT, 42% (20), 20% (21); b) H2, Pd(OH)2, EtOH, 87%.  $DIAD = diisopropylazodicarboxylate, F-PPh_3 = diphenyl-[4-(1H,1H,2H,2H-perfluorodecyl)phenyl]phoselement of the property of$ phine.

considerable experimentation, we identified the Mitsunobu reaction<sup>[1f,15]</sup> as a suitable transformation for fragment coupling of 4 and 5. However, in this particular instance, separation of the triphenylphosphine oxide by-product from the desired products was found to be problematic. Fortunately, use of a fluorous-tagged triphenylphosphine reagent (F-PPh<sub>3</sub>)<sup>[16]</sup> enabled facile product purification after filtration through fluorous silica gel. The desired coupling product was isolated as a separable mixture of diastereoisomers 20 (42%) and 21 (20%). Finally, hydrogenation of 20 and 21 using Pearlman's catalyst afforded silvestrol (1) and its 1" stereoisomer 22. Data for synthetic 1 were confirmed to be identical with those reported for natural (-)-silvestrol<sup>[2]</sup> including <sup>1</sup>H and  $^{13}\text{C NMR}$ , mass, and IR spectra,  $[\alpha]_{\text{D}}$  values, HPLC, and TLC  $R_f$  values.<sup>[17]</sup>

Cyclopenta[b]benzofurans are known protein synthesis inhibitors.[18,19] Accordingly, we compared the relative potencies of silvestrol (1) and its 1" diastereoisomer 22 in an in vitro translation system utilizing a rabbit reticulocyte lysate programmed with firefly luciferase (FLuc) mRNA.[20] Titration of silvestrol (1) revealed an IC<sub>50</sub> value of approximately 0.4 μM, whereas 22 showed an  $IC_{50}$  value of about 2  $\mu M$ (Figure 3a), indicating that 22 is fivefold less active than 1. The relative potency of 1 and 22 was also assessed in vivo by exposing cells to the two compounds, followed by monitoring the incorporation of <sup>35</sup>S-methionine into proteins. The results indicate a 10-fold difference in IC<sub>50</sub> values between silvestrol (1) and 22, with silvestrol being more potent for inhibition of protein synthesis in HeLa cells (Figure 3b).

In conclusion, we have accomplished the enantioselective synthesis of the rocaglate natural product and antitumor agent (-)-silvestrol. The key

> strategy involves an enantioselective dipolar cycloaddition of oxidopyrylium ylides derived from excited-state intramolecular proton transfer (ESIPT) of 3hydroxyflavones using specifically functionalized TADDOL derivatives as chiral Brønsted acids. The unusual 1,4-dioxanyl unit was generated from readily available starting materials using a tandem alkylation/lactonization sequence. Initial biological studies indicate that silvestrol has approximately a 5-10-fold greater activity as an inhibitor of protein synthesis in vivo and in vitro in HeLa cells than its 1"" diastereomer, illustrating the influence of the stereochemistry of the dioxanyl moiety on biological activity.[21] Further studies toward the synthesis of related rocaglamide derivatives and bio-

logical evaluation of silvestrol and related molecules will be reported in due course.

Received: June 20, 2007

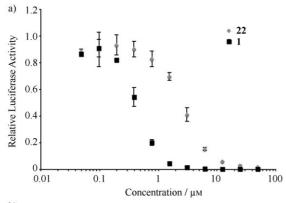
Published online: September 5, 2007

**Keywords:** antitumor agents · cycloaddition · natural products · photochemistry · total synthesis

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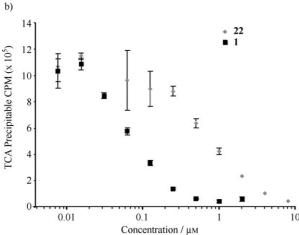


Figure 3. a) Dose-dependent inhibition of translation in vitro by 1 and 22 in rabbit reticulocyte lysates. Firefly luciferase activity of FF/HCV/Ren mRNA in the presence of compound 1 or 22 was normalized to the value obtained in the presence of vehicle (DMSO). b) Dose-dependent inhibition of protein synthesis in vivo in HeLa cells by 1 and 22. TCA: separation of radiolabeled protein from unincorporated 35S-methionine by precipitation. The material was quantified by liquid scintillation counting.

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